© 2005 Adls Data Information BV. All rights reserved.

# Correlation between Seminal Carnitine and Functional Spermatozoal Characteristics in Men with Semen Dysfunction of Various Origins

Michele De Rosa,<sup>1</sup> Bartolomeo Boggia,<sup>1</sup> Biagio Amalfi,<sup>1</sup> Stefano Zarrilli,<sup>1</sup> Angela Vita,<sup>2</sup> Annamaria Colao<sup>1</sup> and Gaetano Lombardi<sup>1</sup>

- 1 Department of Molecular and Clinical Endocrinology and Oncology, University of Naples 'Federico II', Naples, Italy
- 2 Andrology Unit, S. Carlo Hospital, Potenza, Italy

### **Abstract**

Background and objective: L-carnitine is an essential molecule involved in mitochondrial metabolism, controlling the transport of acetyl and acyl groups across the mitochondrial inner membrane. Carnitine and acetylated carnitine (L-acetylcarnitine) are found in high concentrations in the epididymis, where they also act as antioxidants, protecting spermatozoa against damage caused by reactive oxygen species. In this open study we investigated the correlation between seminal carnitine levels and spermatozoal function, and the effect of combined L-carnitine + L-acetylcarnitine therapy, in infertile men.

Patients and methods: 170 infertile men were enrolled in this study. Patients were divided into those with a total sperm motility below the normal WHO range (<50% motility, group 1 [n = 102]) and those with total sperm motility within the normal range ( $\ge50\%$  motility, group 2 [n = 68]). Patients in group 1 were further divided into two groups: those with primary or secondary azoospermia (1B [n = 36]), and those without (1A [n = 66]). Patients in group 1A received L-carnitine 1 g/day and L-acetylcarnitine 500mg twice daily for 6 months. Seminal carnitine levels were measured and correlated with sperm count and motility, eosin test, hypo-osmotic swelling test, acridine orange test for sperm nuclear DNA integrity and sperm kinetics evaluated by computer-assisted sperm analysis in all patients.

**Results:** There was a significant correlation between seminal carnitine concentration and sperm concentration, total sperm count, sperm total motility, rapid forward progression, live sperm count, membrane function, nuclear DNA inte-

grity, capacity for cervical mucus penetration, linearity of spermatic movement, and amplitude of lateral sperm head movement (all p < 0.0001) in the entire study population. In group 1A, there was a significant increase in total motility, live sperm count, membrane integrity and linearity of spermatic movement after 3 and 6 months of L-carnitine/L-acetylcarnitine treatment, and in capacity for cervical mucus penetration after 6 months of treatment, compared with baseline.

Conclusion: Seminal carnitine concentration may be an appropriate marker of sperm and epididymal function. L-carnitine/L-acetylcarnitine treatment may be an effective therapy to improve mainly functional seminal parameters.

L-carnitine (levocarnitine), first isolated from beef muscle in 1905, is an essential molecule involved in mitochondrial metabolism. Free carnitine is obtained from the diet, or produced in the liver and kidney, and only the L-isomer is biologically active in mammals. L-carnitine controls the transport of acetyl and acyl groups across the mitochondrial inner membrane, and its concentration was found to be regulated and stable in humans. The total concentration of carnitine within a cell consists of both free L-carnitine and acylcarnitine esters formed by reaction with fatty acids, catalysed by the enzyme carnitine acyltransferase.

Carnitine and acetylated carnitine are found in high concentrations in the epididymis, [6] and epididymal spermatozoa are extraordinarily rich in carnitine acetyltransferase. The main site of carnitine accumulation is the cauda epididymis, but a high concentration can also be detected in luminal fluid collected by micropuncture from the caput. In the epididymis, the epithelium takes up free L-carnitine from the blood plasma and transports it into the epididymal fluid, where it is concentrated 100-fold. L-carnitine is subsequently taken up by the spermatozoa, where it is accumulated as both free and acetylated (L-acetylcarnitine, levace-carnine) forms. [7]

Free L-carnitine plays a pivotal role in mitochondrial  $\beta$ -oxidation of long-chain fatty acids, [8] which

must be activated by binding to coenzyme A (CoA) to form acyl-CoA. Acyl-CoA molecules are not able to passively cross the internal mitochondrial membrane and must be transported by a specific enzymatic shuttle system, the constituents of which include free carnitine. [9] Carnitine also acts as an antioxidant, protecting spermatozoa against damage caused by reactive oxygen species (ROS).

With the exception of subjects with sperm total motility below the normal range established by the World Health Organization (WHO), it is now well recognised that there is a high correlation between seminal carnitine levels and several functional spermatozoal parameters,[10] indicating that carnitine is important for sperm fecundity. In oligozoospermia, asthenozoospermia and necrospermia, the L-acetylcarnitine concentration in seminal plasma is below that of normal controls.<sup>[7]</sup> In particular, the spermatozoal L-carnitine to L-acetylcarnitine ratio is reduced in asthenospermia.[11] It has been demonstrated that azoospermic patients have a low semen concentration of free carnitine,[12,13] and there is a strong correlation between seminal carnitine levels and the anatomical site of occlusion in patients with obstructive azoospermia.[14]

We investigated the association between seminal carnitine levels and sperm function, and the effect of combined L-carnitine and L-acetylcarnitine treatment, in infertile men.

Table I. Seminal parameters in 170 infertile males with various pathologies divided in groups

EM range mean ±SEM  78 0-70 11.87 1.61  24 0-80 11.67 1.34  29 0-80 24.51 2.24  19 0-75 18.43 1.79  10 0-75 18.43 0.75  11 0-48 0.66 0.94  11 0-85 21.85 2.13		Total group		Group 1			Group 1A	4		Groun 1R	<u>«</u>		Group		
L)         L)         L)         L)         L)           total count         73.58         6.24         0-70         11.87         1.61           otal count         73.58         6.24         0-300         34.99         5.35           otal motility (%)         30.06         1.94         0-80         11.67         1.34           invard         13.76         1.23         0-60         2.55         0.46           sion (%)         43.39         2.28         0-90         24.51         1.79           motic swelling         38.48         2.19         0-75         18.43         1.79           mucus         16.24         1.12         0-36         5.43         0.75           ion capacity         1.22         0.09         0-4.8         0.66         0.94           of spermatic         38.45         2.06         0-75         21.85         2.13           nt         nt         1.23         0.34         0.75         1.36         1.36		1 1	1 1		±SEM	range	mean	±SEM	range	mean	±SEM	range	mean	±SEM	range
tall count 73.58 6.24 0-300 34.99 5.35 otal motility (%) 30.06 1.94 0-80 11.67 1.34 invard 13.76 1.23 0-60 2.55 0.46 sion (%) 43.39 2.28 0-90 24.51 2.24 motic swelling 38.48 2.19 0-75 18.43 1.79 int of the head 11.22 0.09 0-4.8 0.66 0.94 int of the head at of spermatic 38.45 2.06 0-75 21.85 2.13 int orange test (%) 42.31 2.31 0-85 21.86 1.96				11.87	1.61	09-0	18.35	2.09	0.5–60	0	0	0	41.29	2.51	1–70
otal count         73.58         6.24         0-300         34.99         5.35           otal mottlifty (%)         30.06         1.94         0-80         11.67         1.34           inward         13.76         1.23         0-60         2.55         0.46           sion (%)         43.39         2.28         0-90         24.51         1.34           st (%)         43.39         2.28         0-90         24.51         1.79           motic swelling         38.48         2.19         0-75         18.43         1.79           mucus         16.24         1.12         0-36         5.43         0.75           ion capacity         1.22         0.09         0-4.8         0.66         0.94           nt of the head         38.45         2.06         0-75         21.85         2.13           nt         1.36         2.31         0-86         0.94         1.96															
otal motility (%) 30.06 1.94 0–80 11.67 1.34 inward 13.76 1.23 0–60 2.55 0.46 sion (%) 43.39 2.28 0–90 24.51 2.24 motic swelling 38.48 2.19 0–75 18.43 1.79 inn capacity 1.22 0.09 0–4.8 0.66 0.94 int of the head of spermatic 38.45 2.06 0–75 21.85 2.13 in orange test (%) 42.31 2.31 0–86 1.96					5.35	0-240	54.08	7.26	2-240	. 0	0	0	131.50	9.86	1.5-300
otal motility (%) 30.06 1.94 0–80 11.67 1.34 nward 13.76 1.23 0–60 2.55 0.46 sion (%) 43.39 2.28 0–90 24.51 2.24 motic swelling 38.48 2.19 0–75 18.43 1.79 mucus 16.24 1.12 0–36 5.43 0.75 ion capacity 1.22 0.09 0–4.8 0.66 0.94 ant of the head at t															
sion (%)       13.76       1.23       0-60       2.55       0.46         sion (%)       43.39       2.28       0-90       24.51       2.24         motic swelling       38.48       2.19       0-75       18.43       1.79         mucus       16.24       1.12       0-36       5.43       0.75         ion capacity       1.22       0.09       0-4.8       0.66       0.94         int of the head       38.45       2.06       0-75       21.85       2.13         of spermatic       38.45       2.06       0-75       21.85       2.13         int       0range test (%)       42.31       2.31       0-85       21.86       1.96				11.67	1.34	0-40	18.03	1.58	0-40	0	0	0	57.60	0.84	50-80
st (%) st (%) at (3.39  2.28  0–90  24.51  2.24 motic swelling 38.48  2.19  0–75  18.43  1.79 mucus  mucus  16.24  1.12  0–36  5.43  0.75 ion capacity  at of lateral  1.22  0.09  0–4.8  0.66  0.94 ant of the head  of spermatic  38.45  2.06  0–75  21.85  2.13  nt  nt  nt  orange test (%) 43.39  2.28  0–90  24.51  2.31  0–85  21.86  1.96				2.55	0.46	0-50	3.93	0.64	050	0	0	0	30.59	1.44	09-0
st (%) st (%) st (%) st (%) thick swelling (1.28) st (2.28) st (0.90) st (2.51) st (1.79) st (1.12) st (1.	์ (%) บเ									i.					
motic swelling 38.48 2.19 0–75 18.43 1.79 mucus 16.24 1.12 0–36 5.43 0.75 ion capacity  le of lateral 1.22 0.09 0–4.8 0.66 0.94 ant of the head  of spermatic 38.45 2.06 0–75 21.85 2.13 nnt  nt				24.51	2.24	0-65	37.88	2.05	10-65	0	0	0	71.71	1.21	40-90
mucus 16.24 1.12 0–36 5.43 0.75 ion capacity le of lateral 1.22 0.09 0–4.8 0.66 0.94 and of spermatic 38.45 2.06 0–75 21.85 2.13 nnt  orange test (%) 42.31 2.31 0–85 21.86 1.96				18.43	1.79	09-0	28.48	1.81	2-60	0	0	0	96.9	0.63	55–75
ration capacity tude of lateral 1.22 0.09 0-4.8 0.66 0.94 ment of the head ity of spermatic 38.45 2.06 0-75 21.85 2.13 ment ne orange test (%) 42.31 2.31 0-85 21.86 1.96													,		
ration capacity  tude of lateral 1.22 0.09 0–4.8 0.66 0.94  ment of the head  ity of spermatic 38.45 2.06 0–75 21.85 2.13  ment  ne orange test (%) 42.31 2.31 0–85 21.86 1.96				5.43	0.75	0–25	8.39	0.97	0-25	0	0	0	32.44	0.28	27–36
tude of lateral 1.22 0.09 0-4.8 0.66 0.94 ment of the head ity of spermatic 38.45 2.06 0-75 21.85 2.13 ment lith orange test (%) 42.31 2.31 0-85 21.86 1.96	n capacity														
itude of lateral 1.22 0.09 0–4.8 0.66 0.94 iment of the head 38.45 2.06 0–75 21.85 2.13 iment the orange test (%) 42.31 2.31 0–85 21.86 1.96															
ine orange test (%) 42.31 2.31 0–85 21.86 1.96				0.6	0.94	4	1.01	0.13	4-0	0	0	0	2.06	0.09	1.5-4.8
rrity of spermatic 38.45 2.06 0–75 21.85 2.13 rment ine orange test (%) 42.31 2.31 0–85 21.86 1.96	of the head														
38.45 2.06 0–75 21.85 2.13 42.31 2.31 0–85 21.86 1.96		بنعي													
42.31 2.31 0–85 21.86 1.96				21.85	2.13	0-56.2	33.77	2.77	056	0	0	0	63.34	1.06	18.9–75
42.31 2.31 0–85 21.86 1.96									-						
				21.86	1.96	0-65	3.79	1.75	2.65	0	0	0	72.97	1.23	20–85
Carnitine (μmol/L) 266 25.58 2.66- 191 22.79 2.		25.5(			22.79	2.66-	227.70	32.84	7	123.70	19.37	2.66-	379.60	51.31	15.05-
1779 98			1779			7.786			987			421.4			1779

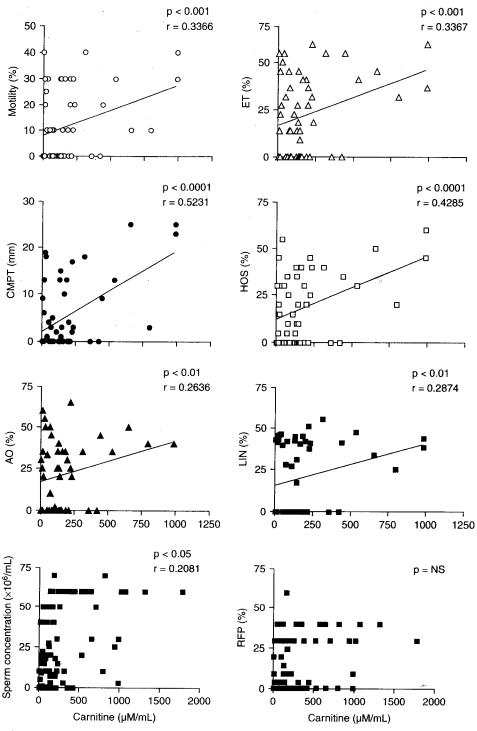


Fig. 1. Correlation analysis between seminal carnitine levels and sperm parameters in patients with low sperm motility (group 1A). AO = acridine orange test; CMPT = cervical mucus penetration test; ET = eosin test; HOS = hypo-osmotic swelling test; LIN = linearity of spermatozoal movement; RFP = rapid forward progression.

## **Patients and Methods**

A total of 170 (duration 1–2 years) infertile patients (aged between 23 and 38 years) without any general illness, presenting with various pathologies

that affect fertility without any female factor, including previous cryptorchidism, testicular atrophy, varicocoele, surgically treated varicocoele, idiopathic asthenozoospermia and asymptomatic bacterio-

**Table II.** Linear regression analysis between carnitine and seminal parameters in 134 male infertile patients (total study group except for males with azoospermia)

Parameter	Mean	±SEM	Range	p-Value	r-Value
Sperm concentration (×106/mL)	29.99	1.91	0.5–70	<0.0001	0.41
Sperm total count (×106)	93.34	6.99	1.5-300	<0.01	0.23
Sperm total motility (%)	38.13	1.93	0-80	<0.001	0.30
Rapid forward progression (%)	17.46	1.4	0–60	<0.01	0.25
Eosin test (%)	55.04	1.88	10–90	<0.001	. 0.31
Hypo-osmotic swelling test (%)	48.82	1.99	5–75	<0.001	0.33
Cervical mucus penetration capacity (mm)	20.6	1.16	0–36	<0.001	0.29
Acridine orange test (%)	53.67	2.0	2–85	<0.01	0.28
Amplitude of lateral movement of the head (μm)	1.55	0.09	04.8	<0.01	0.23
Linearity of spermatic movement	48.78	1.75	0–75	<0.01	0.25

spermia, were enrolled in this study between February 2000 and June 2003. Patients provided written informed consent to be enrolled in the study.

The study group was divided into two groups on the basis of total sperm motility, as defined by the WHO. Group 1 included 102 patients with sperm total motility below the normal range (<50%), while the remaining 68 patients, with total sperm motility within the normal range (≥50%), constituted group 2 (table I). None of the patients had had any treatment affecting spermatogenesis for almost 3 months before study inclusion, including antibiotic treatment for bacteriospermia (washout period).

Within group 1, 66 patients did not have azoospermia and made up group 1A; the remaining 36 patients with primary or secondary azoospermia made up group 1B. Patients in group 1A received L-carnitine 1 g/day and L-acetylcarnitine 500mg twice daily for 6 months. Both drugs were delivered orally and commercially obtained.

All semen analyses were performed according to the 1999 WHO laboratory manual guidelines. The eosin test was used to evaluate live spermatozoa percentage (normal range  $\geq 75\%$ ), the hypo-osmotic swelling test for sperm membrane integrity (normal range  $\geq 60\%$ ), the bovine cervical mucus penetration test for the sperm kinetic index (normal range  $\geq 30$ mm), and the acridine orange test for sperm nuclear DNA integrity (normal range  $\geq 70\%$ ). Sperm

kinetics were evaluated by computer-assisted sperm analysis (CASA) using the Cell Track/S® apparatus (Motion Analysis Corporation, Santa Rosa, CA, USA). This technique was used to determine sperm curvilinear velocity (normal range  $\geq$ 46  $\mu$ m/sec), sperm linear velocity (normal range  $\geq$ 25  $\mu$ m/sec), the linearity of sperm motion (curvilinear velocity divided by linear velocity; normal range  $\geq$ 58%), and the amplitude of lateral sperm head movement (normal range  $\geq$ 1.5 $\mu$ m).

The quantitative evaluation of sperm carnitine levels was performed by spectrophotometry, after enzymatic reaction, according to the method suggested by Marquis and Fritz.<sup>[9]</sup>

Semen analysis was carried out and carnitine levels evaluated in all patients at baseline, and at 3 and 6 months during L-carnitine + L-acetylcarnitine therapy in patients in group 1A.

# Statistical Analysis

A Pearson correlation analysis was performed to investigate the relationship between seminal carnitine concentration and all seminal parameters in group 1A in the entire study population as well as in group 1. Seminal carnitine levels in groups 1 and 2 were compared by t-test to evaluate the differences between the two groups.

In patients treated with L-carnitine + L-acetyl-carnitine, seminal parameters after 3 and 6 months

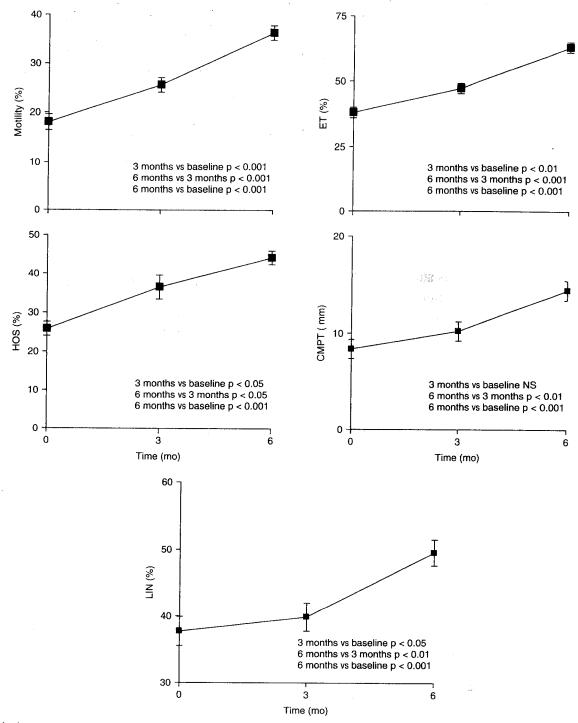


Fig. 2. Seminal parameters at baseline and after 3 and 6 months of treatment in patients with low sperm motility and without azoospermia (group 1A). CMPT = cervical mucus penetration test; ET = eosin test; HOS = hypo-osmotic swelling test; LIN = linearity of spermatozoal movement; NS = not significant.

of therapy were compared with baseline using a Neumann and Keuls test. p-Values <0.05 were considered significant.

### Results

Table I shows the linear regression analysis between seminal parameters and carnitine concentration in the entire study population. There was a significant correlation between seminal carnitine concentration and sperm concentration, total sperm count, sperm total motility, rapid forward progression, live sperm count, membrane function, nuclear DNA integrity, capacity for cervical mucus penetration, linearity of spermatic movement, and amplitude of lateral sperm head movement (all p < 0.0001).

Seminal carnitine levels were significantly lower in group 1 than in group 2 (p < 0.001). In group 1A, out of 66 patients with sperm total motility <50%, there was a significant positive correlation between seminal carnitine concentration and sperm total motility (p < 0.05), live sperm count (p < 0.05), membrane function (p < 0.001), and capacity for cervical mucus penetration (p < 0.0001). The relationship between these seminal parameters and carnitine concentration is shown in figure 1.

In the patients with total sperm motility below the normal range (group 1A), there was a significant increase in total motility, live sperm count, membrane integrity and linearity of spermatic movement after 3 and 6 months of L-carnitine + L-acetylcarnitine treatment, and in capacity for cervical mucus penetration after 6 months of treatment, compared with baseline. Table II and figure 2 summarise results from the Neumann-Keuls test carried out in this group.

### **Discussion**

The hypothesis that, in epididymal spermatozoa, carnitine facilitates fatty acid transport and lipid oxidation enabling the storage of acetyl groups and thereby regulating the level of acetyl—CoA,<sup>[15]</sup> is in concordance with several observations: the presence of significant amounts of free fatty acids in the epididymal plasma,<sup>[16]</sup> the stimulatory effect of carnitine on the incorporation of exogenous palmitic acid into sperm phospholipids and 1,2-diglicerides,<sup>[17]</sup> the ability of spermatozoa to oxidise their own endogenous phospholipids, thereby providing

metabolic energy,<sup>[18,19]</sup> and the presence in the spermatozoa of an appreciable reserve of intracellularlybound acetate after exhaustive washing.<sup>[20]</sup>

Acetylated carnitine is probably the main source of intracellularly bound acetate carried by the epididymal spermatozoa into the ejaculated semen. In human semen, where the concentration of total carnitine as well as acetylated carnitine is much higher in spermatozoa than in seminal plasma, the intracellular level of acetylated carnitine is positively correlated with sperm motility.<sup>[21]</sup> It is highly probable that carnitine and acetylated carnitine are metabolised by spermatozoa in the same way as elsewhere in the body, especially the liver. [22] Intracellular free L-carnitine accumulated by spermatozoa might participate in a buffering role, trapping excess mitochondrial acetyl-CoA as L-acetylcarnitine. This system would protect the activity of pyruvate dehydrogenase, a key enzyme for mitochondrial respiration, which is inhibited by excess acetyl-CoA. Exchanges of free L-carnitine and L-acetylcarnitine between the mitochondrion and the cytoplasm (via a translocase-like enzyme) constitute a carrier system that allows cytoplasmic storage of acetyl groups.

In our study we showed that seminal carnitine concentrations are significantly (p < 0.0001) but weakly correlated with a variety of seminal parameters, regardless of the underlying pathology causing their infertility. Notably, the correlation between carnitine levels and a number of seminal parameters was also significant among the 66 patients with sperm total motility below the normal range (group 1A).

Treatment with L-carnitine in combination with L-acetylcarnitine resulted in significant improvements in several seminal parameters, indicating that carnitine has a beneficial metabolic effect on spermatozoa function, specifically post-testicular sperm maturation, confirming other previous studies.<sup>[23]</sup> Sperm count and sperm morphology, including nu-

clear DNA integrity, were unaffected by L-carnitine + L-acetylcarnitine therapy. As these parameters are a product of intratesticular development, [24-27] this suggests that L-carnitine + L-acetylcarnitine treatment has no effect in this compartment. In spite of the lack of previous double-blind, placebo-controlled studies, [28] Lenzi et al. [29] have demonstrated in a double-blind study on carnitine and L-carnitine treatment of asthenozoospermia that it is effective in increasing sperm motility, especially in groups with lower baseline levels.

These data demonstrate that in a heterogeneous population of infertile men, seminal carnitine concentration plays an important role in spermatogenesis and may be a useful marker of testicular and epididymal function.<sup>[30,31]</sup>

### Conclusion

In conclusion, our data suggest that seminal carnitine concentration is an appropriate marker of sperm and epididymal function. L-carnitine + L-acetylcarnitine treatment may be an effective therapy to improve mainly functional seminal parameters, acting in the post-testicular compartment. Further studies are warranted to confirm the efficacy of L-carnitine + L-acetylcarnitine treatment and to investigate the exact site of action of this combination therapy.

# **Acknowledgements**

We acknowledge the excellent technical assistance provided by Mr Angelo Di Francia. This study had no funding and the authors have no conflicts of interest directly relevant to the content of this study.

### References

- Li B, Lloyd ML, Gudjonsson H, et al. The effect of enteral carnitine administration in humans. Am J Clin Nutr 1992; 55: 838-45
- 2. Casillas ER. Accumulation of carnitine by bovine spermatozoa during maturation in the epididymis. J Biol Chem 1973; 248: 8227-32
- 3. Casillas ER, Chaipayungpan S. The distribution of carnitine and acetylcarnitine in the rabbit epididymis and the carnitine con-

- tent of rabbit spermatozoa during maturation. J Reprod Fertil 1979: 56: 439-44
- 4. Casillas ER, Erickson BJ. Studies on carnitine synthesis in the rat epididymis. J Reprod Fertil 1975; 44: 287-91
- 5. Hinton BT, Snoswell AM, Setchell BP. The concentration of carnitine in the luminal fluid of the testis and epididymis of the rat and some other mammals. J Reprod Fertil 1979; 56: 105-11
- 6. Marquis NR, Fritz IB. Effects of testosterone on the distribution of carnitine, acetylcarnitine, and carnitine acetyltransferase in tissues of the reproductive system of the male rat. J Biol Chem 1965; 240: 2197-200
- Lenzi A, Lombardo F, Gandini L, et al. Metabolism and action of L-carnitine: its possible role in sperm tail function [in Italian]. Arch Ital Urol Nefrol Androl 1992; 64: 187-96
- Jeulin C, Lewin LM. Role of free L-carnitine and acetyl-Lcarnitine in post-gonadal maturation of mammalian spermatozoa. Hum Reprod Undate 1996; 2: 87-102
- Marquis NR, Fritz B. Enzymological determination of free carnitine concentrations in rat tissues. J Lipid Res 1964; 41: 184-7
- Matalliotakis I, Koumantaki Y, Evageliou A, et al. L-carnitine levels in the seminal plasma of fertile and infertile men: correlation with sperm quality. Int J Fertil Womens Med 2000; 45: 236-40
- 11. Golan R, Weissenberg R, Lewin LM. Carnitine and acetylcarnitine in motile and immotile human spermatozoa. Int J Androl 1984; 7: 484-94
- Soffer Y, Shalev DP, Weissenberg R, et al. Survey of carnitine content of human semen using a semiquantitative auxanographic method: decreased semen total carnitine concentration in patients with azoospermia or severe oligozoospermia. Andrologia 1981; 13: 440-4
- 13. Lewin LM, Shalev DP, Weissenberg R, et al. Carnitine and acylcarnitines in semen from azoospermic patients. Fertil Steril 1981; 36: 214-8
- Eigenmann J, Bandhauer K, Tomamichel G. Seminal carnitine concentration in obstructive azoospermia. Eur Urol 1994; 26: 134-6
- 15. Casillas ER, Erickson BJ. The role of carnitine in spermatozoan metabolism: substrate-induced elevations in the acetylation state of carnitine and coenzyme A in bovine and monkey spermatozoa. Biol Reprod 1975; 12: 275-83
- Brooks DE, Hamilton DW, Mallek AH. Carnitine and glycerylphosphorylcholine in the regroductive tract of the male rat. J Reprod Fertil 1974; 36: 141-60
- 17. Hamilton DW, Olson GE. Effects of carnitine on oxygen uptake and utilization of (U-14C)palmitate by ejaculated bull spermatozoa. J Reprod Fertil 1976; 46: 195-202
- 18. Hartree EF, Mann T. Plasmalogen in ram semen, and its role in sperm metabolism. Biochem J 1959; 71: 423-34
- Hartree EF, Mann T. Phospholipids in ram semen: metabolism of plasmalogen and fatty acids. Biochem J 1961; 80: 464-76
- 20. Brooks DE, Mann T. Pyruvate metabolism in boar spermatozoa. J Reprod Fertil 1973; 34: 105-19
- 21. Johansen L, Bohmer T. Motility related to the presence of carnitine/acetylcarnitine in human spermatozoa. Int J Androl 1979; 2: 202-10

- 22. Wolf G. Recent research on carnitine: its relation to lipid metabolism. Cambridge (MA): MIT Press, 1965
- 23. Vicari E, La Vignera S, Calogero AE. Antioxidant treatment with carnitines is effective in infertile patients with prostatovesiculoepididymitis and elevated seminal leukocyte concentrations after treatment with nonsteroidal anti-inflammatory compounds. Fertil Steril 2002; 78: 1203-8
- Costa M, Canale D, Filicori M, et al. L-carnitine in idiopathic asthenozoospermia: a multicenter study. Italian Study Group on Carnitine and Male Infertility. Andrologia 1994; 26: 155-9
- Vitali G, Parente R, Melotti C. Carnitine supplementation in human idiopathic asthenospermia: clinical results. Drugs Exp Clin Res 1995; 21: 157-9
- Frenkel R, McGarry J. Carnitine biosynthesis, metabolism and function [Italian edition]. New York (NY): Academic Press, 1984
- 27. Lenzi A, Lombardo F, Sgro P, et al. Use of carnitine therapy in selected cases of male factor infertility: a double-blind cross-over trial. Fertil Steril 2003; 79: 292-300
- 28. Agarwal A, Said TM. Carnitines and male infertility. Reprod Biomed Online 2004; 8: 376-84

- Lenzi A, Sgro P, Salacone P, et al. A placebo-controlled doubleblind randomized trial of the use of combined l-carnitine and lacetyl-carnitine treatment in men with asthenozoospermia. Fertil Steril 2004; 81: 1578-84
- 30. Lewin LM, Beer R, Lunenfeld B. Epididymis and seminal vesicle as sources of carnitine in human seminal fluid: the clinical significance of the carnitine concentration in human seminal fluid. Fertil Steril 1976; 27: 9-13
- 31. Wetterauer U, Heite HJ. Carnitine in seminal plasma: its significance in diagnostic andrology. Arch Androl 1980; 4: 137-43

Correspondence and offprints: Dr *Michele De Rosa*, Department of Molecular and Clinical Endocrinology and Oncology, University of Naples 'Federico II', Via S. Pansini 5, Naples, 80131, Italy.

E-mail: miderosa@unina.it